

Prevalence, Distribution, and Associated Factors of Surgical Site Infections Linked to Pseudomonas aeruginosa with oprL Virulence Gene in Bauchi, Nigeria

Onvi, T*1², Doko, M. H. I² and Ella, E.E²

¹Department of Community Medicine, Abubakar Tafawa Balewa University, Bauchi, Nigeria; ²Department of Microbiology, Ahmadu Bello University, Zaria, Nigeria

*Corresponding author: hardertt@gmail.com

Article History	Abstract
Received: 11 August 2024 Accepted: 12 September 2024 Published: 24 October 2024	Surgical site infections (SSIs) are a global healthcare challenge. The Pseudomonas aeruginosa is a significant SSI pathogen with the oprL gene conferring its multidrug resistance. Although extensive studies on pathogens in infection control are recommended, a study of this kind has not been conducted in Bauchi, Nigeria. Objectives: This study determined the prevalence and associated factors for SSIs linked with <i>P. aeruginosa</i> with oprL gene. Surgical site swabs (n=250) collected from two hospitals in Bauchi, Nigeria, were cultured on Cetrimide agar to isolate <i>P. aeruginosa</i> . The oprL genes of the isolates were detected through a Polymerase chain Reaction. Chisquare tests, Fisher's exact test, and binary logistic regression analyzed the association between SSIs and other variables. About 2% (5/250) of the swabs were infected with <i>P. aeruginosa</i> with oprL gene. SSI occurrence was statistically associated with postoperative duration (p = 0.01). Prolonged postoperative periods increased the odds of developing SSIs by a factor of 3.81 (O.R.:3.81, p=0.02). No significant associations were found between SSIs and sex (p = 0.65), age (p = 0.09), surgery (p = 0.06), and HIV status (p = 0.31). The prevalence of <i>P. aeruginosa</i> linked to SSIs was low. Prolonged postoperative care significantly increased the risk of SSIs. Future research with longitudinal designs and large samples will monitor additional risk factors of SSIs. Infection control should emphasize post-operative care for SSIs. The integration of molecular confirmation of P. aeruginosa isolates should be routine in SSI management.
	Keywords: Nigeria, Pathogen, Prevalence, Pseudomonas, Surgical Site, Infection License: CC BY 4.0*

How to cite this paper: Onvi, T. et al., 2024 Prevalence, Distribution, and Associated Factors of Surgical Site Infections Linked to Pseudomonas aeruginosa with oprL Virulence Gene in Bauchi, Nigeria. Journal of Public Health and Toxicology Research, 2(2): 86-96.

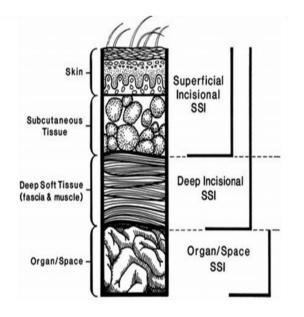
Introduction

A surgical site infection (SSI) is an infection in the part are shown in Figure 1. of the body where surgery takes place. Surgical site Patients with SSIs have purulent discharge at the incision the incision. They are deep incisional when they involve pathogen, the skin's muscle or connective tissue layers. They are Iglewski, & Pesci, 2000). organ/space when they involve structures, organs, or

spaces deep into the incision (Horan et al., 1992). These

infections are defined as those confined to the incisions site or/and at least one of the following: pain, tenderness, and involve structures adjacent to the wounds that were or high temperatures, greater than forty degrees exposed during operation within one month after a Fahrenheit (El Zowalaty & Gyetvai, 2016). In surgical operation or one year after implant surgery immunocompromised patients, with discontinuity in (Berríos-Torres, 2017). SSIs are superficial incisional their epithelial tissue, SSIs are caused by a spectrum of when they involve the skin and subcutaneous tissue of microorganisms including a notable multidrug-resistant Pseudomonas aeruginosa (McKnight,

[•] This work is published open access under the Creative Commons Attribution License 4.0, which permits free reuse, remix, redistribution and transformation provided due credit is given.



Pseudomonas aeruginosa (P. aeruginosa) is a gramnegative bacterium known for its resilience and The SSIs linked to P. aeruginosa vary across antibiotic resistance. It has been linked to several hospital-acquired diseases, including surgical site antibiotic resistance (Mohammad, 2013). This gene SSIs reported the range of the prevalence of P. its reservoir.

introduction of the bacilli from the surgical site (Pollack, by the risk factors of the SSIs. 2000). These three stages are (1) bacterial attachment on The risk factors of SSIs are the same as those of SSIs

polysaccharide slime called biofilm (Brandenburg et al., 2015; Jensen et al., 2017; Maunders & Welch, 2017). (2) local invasion of the whole epidermis tissue and possibly the subcutaneous tissue. This is aided by flagella attachment, biofilms, and extracellular enzymes (Garcia et al., 2018). This stage impairs skin epithelial repair mechanisms and resists phagocytosis by the host immune system and (3) disseminated systemic disease, by blood-borne transfer of the infection, to other distant organs. The virulence factors are crucial in these stages while simultaneously aided by the pathogen's cell-to-cell communication systems called quorum sensing (Lee & Zhang, 2015). Following the invasion of the primary site, there is development of clinical signs of SSI including tenderness and purulent discharge from the surgical site. There may be secondary infections including urinary tract infections (UTIs), otitis externa, meningitis, brain abscess osteomyelitis, endocarditis, and a characteristic skin lesion called Ecthyma gangrenosum (Adhikari et al.,2019).

geographical settings due to host resistance and the pathogenicity of *Pseudomonas* strains. This also affects infections (SSIs) (Klockgether & Tümmler, 2017; Tang the prevalence of P. aeruginosa across settings. The et al., 2017; Paul, 2018). The bacterium is particularly prevalence of SSIs and SSIs associated with P. pathogenic and equipped with various virulence factors aeruginosa vary across settings (Lamichhane et al., including the oprL gene that assists in drug efflux and 2020). A systematic review of the global prevalence of codes the outer membrane proteins of *P. aeruginosa* and *aeruginosa* in SSIs from 4.09% to 26.7% (Reid & Porter, plays an important role in the interaction of the bacterium 1981). Similarly, P. aeruginosa was linked to 13% of with the environment (Wu et al., 2015). These are L- SSIs in a hospital setting in Mumbai, India (Marzoug et peptidoglycan-associated lipoprotein and I-lipoproteins al., 2023). In Brazil, the prevalence of P. aeruginosa outer membrane proteins of *P.aeruginosa*. They alter the linked to SSIs was reported as 11.6% of SSIs (Shah et membrane permeability and utilize the efflux mechanism *al.*,2020). A systematic review of infected wound, skin, to confer antibiotic resistance for *Pseudomonas* species soft tissue, and surgical site infections in Central, and Pseudomonas aeruginosa respectively. These L and Eastern, Southern, and Western Africa, reported the I outer membrane proteins are coded by the oprL and prevalence of P. aeruginosa in infected wound, skin, soft oprI genes respectively. While the oprI is present in all tissue, and surgical site infections as 14% (CI 11% to *Pseudomonas* species, the oprL is specifically present in 18%) (Tuon et al., 2019). The study also found that out P. aeruginosa. Other intrinsic resistance factors encoded of the Gram-negative bacilli isolated, P. aeruginosa was in plasmids are toxA, exoU, exoS, exoY, lasA, lasB, more commonly isolated from the infected wounds oprD, oprL, and oprI. (Peters & Galloway, 1990; (21%, CI 15% to 28%). P. aeruginosa was found to be Galloway, 1991; Choi et al., 2002; Haghi et al., 2018). multidrug-resistant to anti-pseudomonal carbapenems These genes also confer multidrug resistance to the (imipenem or meropenem) in $\geq 20\%$ of isolates (Tuon et pathogen following the invasion of a surgical site from *al.*,2019). In Nigeria, the prevalence of SSI was found to be 14.8% of surgeries (Monk et al., 2024). Out of this, The reservoirs of *P. aeruginosa* include water pools, 4.8% of the Gram-negative bacteria that infected the fruits, flowers, and unsterilized medical and hospital wound were P. aeruginosa (Monk et al., 2024). In instruments and devices. SSIs linked to P. aeruginosa another setting in Nigeria, the prevalence of P. usually occur after some alteration of the normal skin aeruginosa was 19% of the pathogens associated with defense or architecture by the surgical incision. There are SSIs (Abiodun et al., 2014). The variations in the three distinct stages in the pathogenesis of SSIs after the prevalence of SSIs linked to *P. aeruginosa* are affected

compromised epithelial tissues. This is aided by the pilli linked to P. aeruginosa with oprL gene. These risk and flagella of the bacilli. Subsequently, there is factors may be divided into two distinctive categories: colonization of the host tissue with extracellular patient-related (intrinsic) and process/procedural-related (extrinsic) factors (Bucataru *et al.*, 2023). The patient- Methodology related risk factors include factors intrinsic to the Study Design: The approval for the study was obtained patient's health, such as underlying medical conditions from the Bauchi State Health Research Ethical or immunosuppression. Modifiable patient-related risk Committee and the research approval number is factors may include behaviors such as smoking, NREC/12/05/2103/2018/05. The study was a crossalcoholism, or obesity. Smoking, alcoholism, or obesity sectional hospital-based study. It was conducted in two profoundly impact wound healing and immune function selected secondary hospital facilities in the Bauchi local through impairment of oxygen delivery to tissues, government of Bauchi State, in the North-East geomalnutrition, impaired wound closure, and increased political region of Nigeria. The hospitals are New susceptibility of the tissue to infection respectively General Hospital Bayara and Specialist Hospital Bauchi. (Bucataru et al., 2023). The non-modifiable patient- These are shown in Figure 2. related risk factors include age or gender (Bucataru et al., 2023; Trevejo-Nunez et al., 2015). Age and gender affect wound healing through immunity and surgery type respectively. The other category is process- and procedure-related risk factors.

Process- and procedure-related risk factors are extrinsic risk factors associated with the surgical process. They include intraoperative risk factors e.g. asepsis maintenance or the length of the procedures. Others are emergency surgeries, surgical site contamination, extended hospital stays, use of foreign bodies such as transplants, use of a heartimplants and lung machine, blood transfusions, immunosuppressive medication, contamination of the allograft, shaving of t he operative site, ventilation, inadequate sterilization, poor preoperative antibiotic prophylaxis, and poor postoperative wound care and monitoring (Bucataru et al., 2023).

infections like P. The investigation of human aeruginosa and its prevalence within a hospital is essential for the articulation of effective preventive measures. Moreover, the diagnosis of infection prevention techniques requires an understanding of the pathogen's prevalence and distribution (WHO,2017; Pinchera et al., 2022). The World Health Organization, WHO, recommends the prioritization of extensive research on hospital-specific SSIs for infection control and to build national data on Hospital hospital-acquired infections (HAIs) (WHO,2017; Tacconelli et al., 2018; Pinchera et al., 2022). Bauchi in North-East Nigeria has unique challenges in healthcare delivery and a huge burden of SSIs (Olowo-Okere et al., 2019; Onyi et al.,2024a; Onyi et al.,2024b). There is a paucity of studies on the prevalence and distribution of surgeries infected with P. aeruginosa. Most of the strains of the bacilli are multidrug-resistant and have the oprL virulence factor. These strains are associated with SSIs with poor health outcomes including poor wound healing, extended hospital stays, and deaths. The study was an attempt to bridge this research gap. The findings might optimize the management of these SSIs and provide reference data for other related studies. The specific objectives of the study were to investigate the (1) prevalence and (2) factors associated with SSIs linked to P. aeruginosa with the oprL gene.

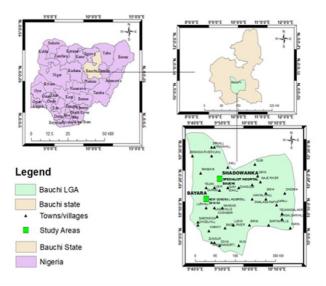


Figure 2. Map of Bauchi State of Nigeria showing the Study area (Onuh, 2019).

Sample Size Determination

The sample size for the study was determined using the Cochrane formula for cross-sectional studies (Cochran, 1963). A prevalence of 19% reported for P. aeruginosa in surgical site infection by Yazidi et al. (2015) was utilized for estimating the minimum sample size. $n=Z^2Pq/L^2$

Where n = number of samples

Z = standard normal deviate at 95% CI = 1.96

P = 19% (Yasidi *et al.*,2015) = 0.19 g = 1 - 0.19 = 0.81

L = precision of 5% (0.05)

 $n = 1.96^2 \ge 0.19 \ge 0.81/0.05^2 = 0.59122224/0.0025 =$ 236.48 ~ 237

With a non-response rate assumed to be 5%, the 237 was multiplied by a factor of 1.05, giving 248.85. This was approximated to 250. A sample size of 250 surgical swabs was utilized for the study. The inclusion criteria for the participants were as recommended by the Centre for Disease Control, CDC. The study included consented patients who had surgeries and were on admission within 30 days of the surgeries; and who had purulent discharge from the surgical wound sites or/and at least had one of the following: pain, tenderness, or high temperature (greater than 40 degrees Fahrenheit (Bucataru et al., 2023). Patients who had antibiotics two weeks before the surgery or who did not consent to the study were excluded.

c. Samples collection, bacterial isolation and identification

Two hundred and fifty surgical site wound swabs were collected from Bauchi Specialist Hospital (205 samples) and New General Hospital Bayara patients (45 samples) within five months from January 2019 to May 2019. A total of two hundred and fifty (250) surgical wound swab samples were collected during wound review or wound dressings at the wards. The swabs were labeled and transported in sterile ice packs to the Abubakar Tafawa Balewa University Teaching Hospital's (ATBUTH's) Microbiology Unit for sample analysis. The sample analysis involved three steps: identification of colonial morphology of culture on Cetrimide agar, Gram staining, and biochemical analysis of isolates (Public Health England,2015). For the phenotypic, there was an identification of culture on Cetrimide agar. The Gram staining of the pure isolates of P. aeruginosa isolates was conducted as described by Bartholomew and Mittwer (1952). The biochemical characterization of the isolates was conducted with Oxidase and Catalase tests. The Molecular confirmation of isolates was with Polymerase Chain Reaction, PCR, with oprL, with an amplicon size of 504, as the target gene. The details of these procedures are highlighted in other study reports (Public Health England, 2015; Tacconelli et al., 2018; Olowo-Okere et al., 2023)

D. Data analysis: The included patients' folders were sought for data extraction. The variables extracted from patients' hospital folders were health facility, ward, age, sex, surgery type, duration in hospital, HIV status, and occupation. Other variables that were included after laboratory analysis were phenotypic identification of colony, Gram staining result, Catalase test, Oxidase test, and molecular confirmation of isolates. The data are recorded in a data extraction sheet and then transferred to Excel Spreadsheet and Statistical Products and Service Solutions (SPSS) version 25 for analysis. A p-value less than 0.05 was considered statistically significant for the Chi-square tests, Fisher's Exact tests, and binary logistic regression analysis.

Results

Demographic characteristics of respondents

The demographic variables of the patients with SSIs linked to *P. aeruginosa* are shown in Table 1

 Table 1: Demographic characteristics of respondents

Variable	No. Test	No. Positi	% positi	χ^2	d f	p- val
s	ed	ve	ve			ue
Sex						

					-		
Female	107		3	2.80	0.6 2	1	0.4 3
Male	143		2	1.40			5
Age							
1-10	10	1		10.00	10. 01	5	0.0 8
11-20	62	1		1.61			
21-30	89	0		0.00			
31-40	61	1		1.63			
41-50	19	1		5.26			
51-60	9	1		11.11			
Educatio							
n status							
No formal educatio n	119	1		0.84	3.5 4	3	0.3 2
Primary	88	2		2.27			
Secondar y	31	1		3.22			
Tertiary	12	1		8.33			
Occupat							
ion							
Unemplo yed	9	0		0.00	9.1 2	1 0	0.5 2
Domestic work	43	1		2.32			
Student	46	1		2.17			
Manual labor	9	0		0.00			
Trading	85	0		0.00			
Hotel manage ment	1	0		0.00			
Professio nal soccer	1	0		0.00			
player Public	3	0		0.00			
relations	5	U		0.00			
Civil service	14	0		0.00			
Set vice $\sum_{i=1}^{n} \gamma^{2} = ch$		 	- 4 1	C 1	1		

Key: χ^2 = chi-squared test; df = degree of freedom; p-value ≤ 0.05 is statistically significant

Data on demographic features are shown in Table 1. It shows the variables - occupation, age, sex, and educational status. Additionally, it shows the number of tests, the number of positive cases, the percentage of positive cases, the degrees of freedom, p-values, and the chi-square statistics of each variable. The subunits of each variable are shown in the rows For the "Sex" variable, the range of values includes "Female" and "Male." The highest number tested for this variable is 143 for males, with 2 positive cases and a percentage positive of 1.40%. The lowest number tested is 107 for females, with 3 positive cases and a percentage positive of 2.80%. The chi-square statistic is 0.62 with 1 degree of freedom, and the p-value is 0.43.

For the "Age" variable, the range of values includes different age groups. The highest number tested is 89 for the 21-30 age group, with 0 positive cases. The lowest number tested is 9 for the 51-60 age group, with 1 positive case and % percentage positive of 11.11%. The chi-squared statistic varies for different age groups is 10.01, with degrees of freedom of 5, and a p-value of 0.08.

For the "Education Status" variable, the range of values education," includes "No formal "Primary," "Secondary," and "Tertiary." The highest number tested is 119 for individuals with no formal education, with 1 positive case and a percentage positive of 0.84%. The lowest number tested is 12 for individuals with tertiary education, with 1 positive case and a percentage positive of 8.33%. The chi-square statistic is 3.54, with degrees of freedom of 3, and a p-value of 0.32

For the "Occupation" variable, the range of values includes a variety of occupational categories such as Unemployed, Domestic work, Student, Manual labor, Trading, Hotel management, Professional Soccer player, Public relations, and Civil service. The highest number tested is 85 for individuals engaged in trading, with 0 positive cases, resulting in a percentage positive of 0.00%. The lowest number tested is 1 for individuals in several categories such as Hotel management, Professional Soccer player, Public relations, and Civil service, with 0 positive cases for each of these categories. The chi-square statistic is 9.12. The degree of Key: χ^2 = chi-squared test; df=degree of freedom; pfreedom is 10, and the corresponding p-value is 0.52.

Prevalence of positive isolates

identified as P. aeruginosa had a colony morphology in two different hospitals-Specialist Hospital Bauchi and characterized by a blue-green smooth colony with New General Hospital Bayara. In Specialist Hospital smooth edges and an elevated appearance. The colonies Bauchi, out of 205 swabs that were analyzed, there were had the smell of 'grapefruit'. On Gram staining, they 5 confirmed positive isolates, accounting for 2.4% of were Gram-negative rods. They were also Oxidase and positive isolates. Forty-five (45) swabs from New isolates Five Catalase positive. aeruginosa positive. Moreover, all five (5) isolates (A- isolates. This data suggests that Specialist Hospital E) showed the oprL gene with an amplicon size of Bauchi had a higher percentage of positive isolates 504bp. This is shown in Figure 2. Therefore, this study compared to New General Hospital Bayara. The γ^2 value found a prevalence of 2% (5/250) of Pseudomonas is 1.12, with 1 degree of freedom, and a p-value of 0.29, infection associated with surgical site infections.

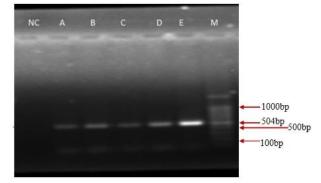


Figure 3: Gel electrophoresis of oprL gene with amplicon size of 504 bp

Key:

Lane NC = Negative control

Lanes A-E = Samples

Lane M = 100bp molecular marker

Prevalence of positive isolates at the facilities

The Prevalence of positive isolates at the facilities is shown in Table 2.

Table 2: Prevalence of positive isolates at the facilities.

Hospit al Faciliti es	No of swabs analyz ed	No positive isolated confirm ed	% of positi ve isolate s	χ2	d f	p- valu e
Speciali st Hospita 1 Bauchi	205	5	2.4	1.1 2	1	0.29
New General Hospita I Bayara	45	0	0.0			

value ≤ 0.05 is statistically significant

Five isolates that were analyzed and phenotypically Table 2 shows the analysis of swabs and positive isolates were *P*. General Hospital Bayara were analyzed with no positive indicating no statistical significance.

Factors associated with surgeries infected with Pseudomonas aeruginosa

The Factors associated with surgeries infected with P. aeruginosa are shown in Table 3.

 Table 3 Factors associated with surgeries infected with P. aeruginosa

Variables	No. Test	No. Posit	% posit	Fish er's	P val
	ed	ive	ive	Exac t	ue
Sex					
Female	107	3	2.80		0.6 5
Male	143	2	1.40		
Age					
1-10	10	1	10.0 0	7.73	0.0 9
11-20	62	1	1.61		
21-30	89	0	0.00		
31-40	61	1	1.63		
41-50	19	1	5.26		
51-60	9	1	11.1 1		
Surgery type					
Cesarean section	105	2	1.90	19.4 2	0.0 6
Laparotomy	38	1	2.63		
Sutured	20	0	0.00		
Laceration					
(upper					
limb/arm)	1		0.00		
Amputation of Right toe	1	0	0.00		
Inguinal/Herni orrhaphy	19	0	0.00		
Sutured Breast	6	1	16.6		
lumpectomy			7		
Incisional hernia	3	0	0.00		
Appendectomy	5	0	0.00		
Amputation (Below the knee)	28	0	0.00		
Sutured laceration scalp	18	0	0.00		
Amputation below elbow	1	1	100. 00		
Sutured	6	0	0.00		
laceration (left foot)					
Post-operative			<u> </u>		
duration					
1-5	206	2	0.97	11.3 9*	0.0
6-10	28	1	3.57	,	1
11-15	16	2	12.5		
			0		

Research	article
----------	---------

HIV status				
Positive	8	1	12.5	0.3
			0	1
Negative	242	4	1.65	

Key: Fisher's Exact test is utilized when at least 80% observed frequency is less than 5. Exact significance \leq 0.05 is statistically significant (*). All values approximated to 2 decimal places

Table 3 shows the risk factors associated with Surgical Site Infections (SSIs) linked to *P. aeruginosa* are presented. The variables are age, sex, surgery type, post-operative duration, and HIV status. The number of tests, positive cases, percentage of positive cases, Fisher's exact Value, and Exact significance p value (2-sided) were detailed for each variable.

In the analysis of SSIs, the "Sex" variable was examined. It shows that out of 143 males tested, 2 cases were positive, yielding a percentage positive of 1.40%. Similarly, out of 107 females tested, 3 cases were positive, resulting in a percentage positive of 2.80%. Fisher's Exact Test yielded a value of 0.65, with an Exact significance (2-sided) of 0.25, indicating that the difference in SSIs between the sexes is not statistically significant.

Age groups were also investigated, showing varying percentages of positive cases across different ranges. Notably, the age groups 1-10 and 51-60 demonstrated higher percentages of SSIs, ranging from 0% to 11.11%. Despite a Fisher's Exact value of 7.73, the Exact significance (2-sided) of 0.09 suggests that the age groups are not significantly associated with SSIs.

Analysis of surgery types revealed that patients undergoing amputation below the elbow showed the highest percentage of SSIs at 100%. However, Fisher's Exact value of 19.42 and an Exact significance of 0.06 indicated an insignificant association between the type of surgery and SSIs.

Post-operative duration showed that longer durations were associated with higher percentages of SSIs, with a significant p-value of 0.01. The highest percentage of 12.50% was recorded for post-operative duration of 11-15 days.

Additionally, for HIV status, a higher percentage of SSIs was observed in HIV-positive patients (12.50%) compared to HIV-negative patients (1.65%). However, the Exact significance (2-sided) of 0.31 indicated that this difference was not statistically significant.

The SSI data was further regressed against some of the variables that are known risk factors of SSIs. The binary logistic regression analysis involved two steps- bivariate and multivariate regression analysis. This bivariate analysis is shown in Table 4 while the multivariate analysis is shown in Table 5.

Table4	SSI	risk	factors:	Bivariate	regression
analysis					

analys	15					
Variab	les	В	S.E.	Wald	df	Sig.
Age	0.62	0.38	2.76	1	0.10	
Consta	nt	-6.11	1.56	15.38	1	0.00
Sex	-0.76	0.92	0.68	1	0.41	
Consta	nt	-2.76	1.37	4.04	1	0.04
Surger	y0.04	0.12	0.12	1	0.73	
Consta	nt	-4.06	0.68	35.53	1	0.00
Post-O	peration	Duration	n (Days)	1.38	0.52	6.90
	1	0.01*				
Consta	nt	-6.02	1.12	28.84	1	0.00
HIV St	tatus	-1.21	1.14	1.12	1	0.29
Consta	nt	-1.62	2.12	0.59	1	0.44

Table 4 shows the results of a binary logistic regression analysis on surgical site infection and its potential risk factors. The variables examined are age, sex, surgery type, post-operative duration, and HIV status. These are individual bivariate analyses that have been combined in the same table for the sake of simplicity.

Age: The variable age yielded a coefficient of 0.62 with a standard error (S.E.) of 0.38. The Wald statistic was 2.76 with 1 degree of freedom (df), and the significance level (Sig.) was 0.10.

Sex: The coefficient for sex was -0.76 with a standard error of 0.92. The Wald test yielded a statistic of 0.68 with 1 degree of freedom, and the significance level was 0.41.

Surgery: The variable surgery showed a coefficient of 0.04 with a standard error of 0.12. The Wald statistic was 0.12 with 1 degree of freedom, and the significance level was 0.73.

Post-Operative Duration: This variable had a coefficient of 1.38 and a standard error of 0.52. The Wald statistic was 6.90 with 1 degree of freedom, and the significance level was 0.01.

HIV Status: The variable HIV status yielded a coefficient of -1.21 with a standard error of 1.14. The Wald statistic was 1.12 with 1 degree of freedom, and the significance level was 0.29.

Based on the coefficients and significance levels, the post-operative duration exhibited a statistically significant association with the occurrence of surgical site infection (Sig. = 0.01). Age also showed a trend towards significance (Sig. = 0.10), suggesting a potential relationship with the SSIs.

The variables were further analyzed with multivariate analysis shown in Table 5.

 Table 5: SSI risk factors:
 Multivariate regression

 analysis
 Provide the second secon

Variables	В	S.E	Wal	d	Sig.	Exp(B
		•	d	f)
Surgery	0.0	0.1	0.00	1	0.9	1.01
	1	7			7	
Post-	1.3	0.5	5.41	1	0.0	3.82
Operatio	4	8			2	
n						
duration						
Sex	-	1.2	0.00	1	0.9	0.97
	0.0	5			8	
	3					
Age	0.4	0.3	1.30	1	0.2	1.54
	3	8			6	
HIV	-	1.3	0.80	1	0.3	0.30
Status	1.2	6			7	
	1					
Constant	-	4.2	1.50	1	0.2	0.01
	5.1	2			2	
	7					

From Table 5, none of the variables -surgery, sex, age, and HIV status- have a statistically significant effect on the outcome, as indicated by their higher p-values and odds ratios around 1. However, "Post-Operation Duration" has a statistically significant effect on the outcome, as indicated by a p-value that is less than 0.05(Sig. = 0.02) and an odds ratio of 3.82, suggesting that for each unit increase in post-operative duration, the odds of the having SSIs increase by a factor of 3.82, holding other variables constant. It is important to note that other factors did not confound post-operation duration during the multivariate analysis

Discussion

The study found that the prevalence rate of *P. aeruginosa* associated with SSIs was 2%. This finding indicates a relatively low occurrence of this pathogen in the studied population. It suggests that P. aeruginosa is not a dominant pathogen in linked to the SSIs in this particular study. Other pathogens like Staphylococcus aureus, E. *Coli*, etc. may have been implicated in the bacteriology of the SSIs (Abiodun et al., 2014; Yasidi et al., 2015). Factors such as good wound care practices, patient demographics, effective sterilization practices, and good infection control measures in the healthcare facility environment may have contributed to this low prevalence. (Kaplan et al., 2003; WHO, 2018). The prevalence rate observed in this study differs from those reported in other studies. For example, it is lower than the 4.8% obtained in a hospital facility in Nigeria in 2023. Additionally, it is significantly lower than the 13% reported in India (Marzoug et al., 2023), 11.6% reported in Brazil (Shah et al., 2020), and 14% reported in African regions (Tuon et al., 2019). These variations in prevalence rates may be attributed to differences in wound care practices, patient demographics, hospital

among geographic regions and the immune statuses of site infection, and for identifying immediate risk factors individuals (Kaplan et al., 2003; WHO, 2018). The associated with infections (Mann, 2003). It is also reported prevalence at Specialist Hospital Bauchi and commonly used in epidemiological research to assess the New General Hospital Bayara were 2.4% and 0.0% burden of disease within a population (Mann, 2003). respectively. The variations in the prevalence may be due This design allowed for the analysis of data regarding the to geographical location and patient load. While prevalence of SSIs and provided valuable insights into Specialist Hospital Bauchi is located in the urban city of the factors associated with SSIs linked to *P.aeruginosa*. Bauchi metropolis with more patient load, New General The combined use of culture methods and molecular Hospital Bayara is located in a rural area with a lower techniques enhanced the reliability of the detection of P. patient load.

The study found that SSIs caused by P. aeruginosa are allowed for the growth and initial identification of the significantly associated with the duration of post- bacteria. Then, the molecular technique confirmed the operative care. The binary regression analysis found that presence of oprL resistance genes associated with the for each unit increase in post-operative duration, the odds pathogen, ensuring precise identification (Anuj et of having SSIs caused by P. aeruginosa increased by a al., 2009; Deschaght et al., 2011). Many studies have factor of 3.82, controlling for other variables. The SSIs similarly utilized multi-step identification of P. caused by multidrug-resistant P. aeruginosa require aeruginosa to increase the specificity or identification of extended hospital stays for effective treatment, often pathogens (Anuj et al., 2009; Deschaght et al., 2011) involving long-term use of potent antibiotics, repeated However, the study had some limitations. It was limited surgical debridement, wound care, and sometimes by the cross-sectional design. The cross-sectional design revision surgeries, all contributing to extended hospital does not provide information on the trend or pattern of stays and increased risk of additional complications surgical site infections (SSIs) associated with P. (Rogers et al., 2018). Comparable to the findings in this aeruginosa. A longitudinal study design would track study, Monk et al. (2024) found that patients with P. changes over time and establish temporal relationships aeruginosa SSIs had significantly longer hospital stays between SSIs and P. aeruginosa. The low prevalence compared to those with infections caused by other found in this study suggests that a huge portion of the pathogens. This finding underscores the need for optimal pathogens implicated in the SSIs are not *P. aeruginosa*. surgical site wound care to minimize morbidity However, the study scope was limited to P. aeruginosa associated with wound care and minimize the duration of and was unable to identify other pathogens possibly admission and cost.

Interestingly, this study did not find any statistical been simultaneously utilized to identify these other association between SSI and the type of surgery, age, pathogens to elaborate on the bacteriology of the SSIs. sex, or HIV status. Further analysis showed that none of Moreover, the PCR technique utilized only detected the these variables individually or collectively predicted the *oprL* resistance gene. While the *oprL* gene is a notable likelihood of developing SSI in the presence of other intrinsic resistance gene implicated in the multidrug variables. This finding is contrary to existing knowledge resistance of *P. aeruginosa*, other resistance genes are that suggests these variables are known risk factors for integral in the overall multidrug resistance of P. the development of SSIs linked to P. aeruginosa aeruginosa. (Peters & Galloway, 1990; Galloway, 1991; (Bucataru et al., 2023). The extensive deep tissue Choi et al., 2002; Haghi et al., 2018). Future studies disruption and exposure during these surgeries create a could investigate these virulence factors through conducive environment for colonization by P. multiplex PCR techniques (Anuj et al., 2009). This will aeruginosa (dela Merced et al., 2021; Tornero et enhance a broad understanding of the multidrug al.,2018) Sex is a risk factor for SSIs and is related to resistance of P. aeruginosa associated with SSIs. The surgery type and frequency of surgeries which may vary study did not investigate many other risk factors of SSIs across males and females (Bucataru et al., 2023). HIV associated with P. aeruginosa. These include obesity, and old age are normally associated with a weakened alcoholism, surgical techniques, and duration of immune system, increasing susceptibility to infections, surgeries (Bucataru et al., 2023). This incomplete data including SSIs. However, sample size, random error, and on these variables has the potential of omitting crucial uncontrolled confounders like obesity, alcoholism, and factors that contribute to the SSIs (Deschaght et al., 2011) diabetes may have been a reason for this insignificant. The findings of the study have significant implications association of SSIs and these variables in this study for future research. Future studies should utilize a larger (Penn State University, n.d.).

The study utilized a cross-sectional design, which The studies should investigate more risk factors involved the observation of a defined population at a associated with SSIs. Longitudinal study design should single point in time or over a short period (Mann, 2003). be also utilized to track the trends of SSIs linked to P. This type of study is beneficial for analyzing data

environments, and the gene diversity of *P. aeruginosa* regarding the prevalence of an outcome, such as surgical aeruginosa linked to SSIs. First, the culture methods

> implicated in the SSIs. Other culture media could have sample size to investigate SSIs linked to P. aeruginosa.

Health policies should prioritize stringent infection control measures, especially in post-operative care. Such measures should be conducted with updated surgical wound care guidelines. In the monitoring of SSIs associated with P. aeruginosa, molecular techniques should be routine investigations.

Conclusion

The study reported a 2.0% prevalence of surgical site infections associated with Pseudomonas aeruginosa that Bucataru, A., Balasoiu, M., Ghenea, A. E., Zlatian, O. M., were confirmed with oprL target gene. Post-operative duration increased the odds of developing SSIs associated with P. aeruginosa infection. Further studies with larger sample sizes should utilize longitudinal design to and possibly multiplex PCR techniques to detect and monitor Pseudomonas aeruginosa associated with SSIs.Infection control of SSIs should emphasize postoperative care. These will optimize the treatment outcomes and patients' health.

Acknowledgments

The authors acknowledge all the participants who provided the information and the research assistants who collected the data for this research.

Funding.

The authors have no affiliation with any organization with a direct or indirect financial interest in the subject matter discussed in the manuscript.

Competing Interest

This manuscript has not been submitted to, nor is it under review at, another Journal or other publishing venue. References

- Abiodun, A., Ogunjobi, A. A., & Oluwatosin, O. M. (2014). Multidrug resistance and virulence genes in Escherichia coli and Pseudomonas aeruginosa isolated from diabetic foot infections. African Journal of Clinical and Experimental Microbiology, 15(2), 64-72.
- Adhikari, S., Khadka, S., Sapkota, S., et al. (2019). Prevalence and antibiograms of uropathogens from the suspected cases of urinary tract infections in Bharatpur Hospital, Nepal. Journal of College of Medical Sciences-Nepal, 15(4), 260-266.
- Anuj, S. N., Whiley, D. M., Kidd, T. J., Bell, S. C., Haghi, F., Zeighami, H., Monazami, A., Toutouchi, F., Wainwright, C. E., Nissen, M. D., & Sloots, T. P. (2009). Identification of Pseudomonas aeruginosa by a duplex real-time polymerase chain reaction assay targeting the ecfX and the gyrB genes. Diagnostic Microbiology and Infectious Disease, 63, 127-131.
- Bartholomew, J. W., & Mittwer, T. (1952). The gram stain. Bacteriological reviews, 16(1), 1-29.

- Berríos-Torres, S. I. (2017). Centers for Disease Control and Prevention Guideline for the Prevention of Surgical Site Infection. JAMA Surgery, 152(8), 784.
- Brandenburg, K. S., Calderon, D. F., Kierski, P. R., Brown, A. L., Shah, N. M., Abbott, N. L., ... & Czuprynski, C. J. (2015). Inhibition of Pseudomonas aeruginosa biofilm formation on wound dressings. Wound Repair and Regeneration, 23(6), 842-854. https://doi.org/10.1111/wrr.12365
- Vulcanescu, D. D., Horhat, F. G., ... & Mogoanta, S. S. (2023). Factors contributing to surgical site infections: A comprehensive systematic review of etiology and risk factors. Clin Pract, 14(1), 52-68. https://doi.org/10.3390/clinpract14010006
- Choi, J. Y., Sifri, C. D., Goumnerov, B. C., Rahme, L. G., Ausubel, F. M., & Calderwood, S. B. (2002). Identification of virulence genes in a pathogenic strain of Pseudomonas aeruginosa by representational difference analysis. Journal of Bacteriology, 184(4), https://doi.org/10.1128/JB.184.4.952-952-961. 961.2002
- Cochran, W. G. (1963). Sampling techniques (2nd ed.). New York: John Wiley and Sons Inc.
- dela Merced, P. A., Nadler, E. P., & Hamdy, R. F. (2021). Surgical site infections (SSI) - Prophylaxis and management. In S. T. Verghese & T. D. Kane (Eds.), Anesthetic management in pediatric general surgery. Springer, Cham. https://doi.org/10.1007/978-3-030-72551-8 7
- Deschaght, P., Van Daele, S., De Baets, F., & Vaneechoutte, M. (2011). PCR and the detection of Pseudomonas aeruginosa in respiratory samples of CF patients: A literature review. Journal of Cystic Fibrosis, 10(5), 293-297. https://doi.org/10.1016/j.jcf.2011.05.004
- El Zowalaty, M. E., & Gyetvai, B. (2016). Effectiveness of Antipseudomonal Antibiotics and Mechanisms of Multidrug Resistance in Pseudomonas aeruginosa. Polish Journal of Microbiology, 65(1), 23-32.
- Galloway, D. R. (1991). Pseudomonas aeruginosa elastase and elastolysis revisited: Recent developments. Molecular *Microbiology*, 5(10), 2315-2321. https://doi.org/10.1111/j.1365-2958.1991.tb02076.x
- Garcia, M., Morello, E., Garnier, J., Barrault, C., Garnier, M., & Burucoa, C. (2018). Pseudomonas aeruginosa flagellum is critical for invasion, cutaneous persistence and induction of inflammatory response of skin epidermis. Virulence, 9(1), 1163-1175. https://doi.org/10.1080/21505594.2018.1480830
- Nazaralian, S., & Naderi, G. (2018). Diversity of virulence genes in multidrug-resistant Pseudomonas aeruginosa isolated from burn wound infections. Microbial Pathogenesis, 115. 251-256. https://doi.org/10.1016/j.micpath.2017.12.052
- Horan, T. C., Gaynes, R. P., Martone, W. J., Jarvis, W. R., & Emori, T. G. (1992). CDC definitions of nosocomial

definitions of surgical wound infections. Infection Control & Hospital Epidemiology, 13(10), 606-608.

- Jensen, L. K., Johansen, A. S. B., & Jensen, H. E. (2017). Porcine models of biofilm infections with focus on https://doi.org/10.3389/fmicb.2017.01961
- Microbiology of wound infection after cesarean section in a Jordanian hospital. East Mediterr Health J., 9(5/6), 1069-1075.
- Kaye, K. S., Pogue, J. M., Tran, T. B., Nation, R. L., & Li, J. (2019). Agents of last resort: Polymyxin resistance. Infectious Disease Clinics of North America, 33(4), 835-851. https://doi.org/10.1016/j.idc.2019.08.010
- Klockgether, J., & Tümmler, B. (2017). Recent advances in understanding Pseudomonas aeruginosa as a pathogen. F1000Research, 6, 1261. https://doi.org/10.12688/f1000research.10506.1
- Lamichhane, A., Sapkota, S., Khadka, S., et al. (2020). of lower respiratory tract infection in Bharatpur Hospital, Nepal. Anti-Infective Agents, 18.
- Lee, J., & Zhang, L. (2015). The hierarchy quorum sensing 6, 26-41. https://doi.org/10.1007/s13238-014-0100-x
- Mann, C. J. (2003). Observational research methods. Research design II: Cohort, cross-sectional and case-control studies. Emerg Med J., 20(1),54-60. https://doi.org/10.1136/emj.20.1.54
- Marzoug, O. A., Anees, A., & Malik, E. M. (2023). Pinchera, B., Buonomo, A. R., Schiano Moriello, N., Scotto, Assessment of risk factors associated with surgical site infection following abdominal surgery: A systematic review. BMJ Surgery, Interventions, & Health Technologies, 5(1), https://doi.org/10.1136/bmjsit-2023-000182
- & Welch, M. Maunders, Е., (2017). Matrix exopolysaccharides; the sticky side of biofilm https://doi.org/10.1093/femsle/fnx120
- McKnight, S. L., Iglewski, B. H., & Pesci, E. C. (2000). The Pseudomonas quinolone signal regulates rhl quorum sensing in Pseudomonas aeruginosa. Journal of Bacteriology, 182(10), 2702-2708. https://doi.org/10.1128/JB.182.10.2702-2708.2000
- Mohammad, H. H. (2013). Phenotypic Investigation for Virulence Factors of Pyocine Producing Pseudomonas aeruginosa Isolated from Burn Wounds, Iraq. Reid, T. M., & Porter, I. A. (1981). An outbreak of otitis International Journal of Scientific & Engineering Research, 4(7), 2114-2121.
- Monk, E. J. M., Jones, T. P. W., Bongomin, F., Kibone, W., Nsubuga, Y., Ssewante, N., Muleya, I., Nsenga, L., Rogers, B. A., Sidjabat, H. E., & Paterson, D. L. (2018). Rao, V. B., & van Zandvoort, K. (2024). Antimicrobial resistance in bacterial wound, skin, soft tissue, and surgical site infections in Central, Eastern, Southern, and Western Africa: A systematic review and metaanalysis. PLOS Global Public Health, 4(4), e0003077. Shah, https://doi.org/10.1371/journal.pgph.0003077
- Nguyen, T., Kave, K. S., & Ho, D. (2020). Comprehensive data collection for accurate risk assessment in SSIs. American Journal of Infection Control, 48(6), 669-675. https://doi.org/10.1016/j.ajic.2020.01.017

- surgical site infections, 1992: A modification of CDC Olowo-Okere, A., Ibrahim, Y., Olayinka, B. O., & Ehinmidu, J. O. (2019). Epidemiology of surgical site infections in Nigeria: A systematic review and meta-analysis. The Nigerian Postgraduate Medical Journal, 26(3), 143-151. https://doi.org/10.4103/npmj.npmj_72_
- pathomorphology. Frontiers in Microbiology, 8, 1961. Onuh, C. O. (2019). Site location map: Shadowanka and Bayara areas of Bauchi, Nigeria.
- Kaplan, N., Smadi, A., Al-Taani, M., & El-Qudah, M. (2003). Onyi, T., Doko, H., & Ella, E. (2024). Molecular detection of oprL gene in Pseudomonas aeruginosa associated with surgical site infections in Bauchi, Nigeria. Microbes and Infectious Diseases. https://doi.org/10.21608/mid.2024.261508.1750
 - Onyi, T., Doko, H., & Ella, E. (2024). The antibiogram of multidrug-resistant Pseudomonas aeruginosa with oprL gene associated with surgical site infections in Bauchi, Nigeria. Microbes and Infectious Diseases. https://doi.org/10.21608/mid.2024.261574.1752
 - Paul, J. P. (2018). Pseudomonas aeruginosa. In Principles and Practice of Pediatric Infectious Diseases (pp. 866-870.e1).
 - Incidence of ESBL-producing Gram-negative bacteria Penn State University. (n.d.). Lesson 8 Bias, confounding, random error, and effect modification. Retrieved from https://online.stat.psu.edu/stat507/book/export/html/7 62
 - network in Pseudomonas aeruginosa. Protein & Cell, Peters, J. E., & Galloway, D. R. (1990). Purification and characterization of an active fragment of the LasA protein from Pseudomonas aeruginosa: enhancement of elastase activity. Journal of Bacteriology, 172(5), 2236-2240. https://doi.org/10.1128/jb.172.5.2236-2240.1990
 - R., Villari, R., & Gentile, I. (2022). Update on the management of surgical site infections. Antibiotics, 11, 1608.
 - e000182. Pollack, M. (2000). Pseudomonas aeruginosa. In G. L. Mandell, J. E. Bennett, & R. Dolin (Eds.), Principles and Practice of Infectious Diseases (5th ed., pp. 2310-2327). Churchill Livingstone.
 - formation. FEMS Microbiology Letters, 364(15). Public Health England. (2015). Identification of Pseudomonas species and other Non-Glucose Fermenters. UK Standards for Microbiology Investigations, ID 17, Issue 3. Retrieved from [https://www.gov.uk/ukstandards-for-microbiology-investigations-smiquality-and-consistency-in-clinicallaboratories](https://www.gov.uk/uk-standards-formicrobiology-investigations-smi-quality-andconsistency-in-clinical-laboratories).
 - externa in competitive swimmers due to Pseudomonas aeruginosa. Journal of Hygiene, 86(3), 357-362. https://doi.org/10.1017/s0022172400069114
 - Escherichia coli O25b-ST131: A pandemic multiresistant community-associated strain. The Journal of Antimicrobial Chemotherapy, 73(1), 173-181. https://doi.org/10.1093/jac/dkx430
 - S., Singhal, T., Naik, R., & Thakkar, P. (2020). Predominance of Multidrug-Resistant Gram-Negative Organisms as Cause of Surgical Site Infections at a Private Tertiary Care Hospital in Mumbai, India. Indian Journal of Medical Microbiology, 38, 344-350

- Tacconelli, E., Carrara, E., Savoldi, A., Harbarth, S., Mendelson, M., Monnet, D. L., ... & the WHO Pathogens Priority List Working Group. (2018). Discovery, research, and development of new antibiotics: The WHO priority list of antibioticresistant bacteria and tuberculosis. *Lancet Infectious Diseases*, 18, 318-327. <u>https://doi.org/10.1016/S1473-3099(17)30753-3</u>
- Tang, Y., Ali, Z., Zou, J., Jin, G., Zhu, J., Yang, J., & Dai, J. (2017). Detection methods for *Pseudomonas aeruginosa:* history and future perspective. *RSC Advances*, 7, 51789-51800. <u>https://doi.org/10.1039/c7ra09064a</u>
- Tornero, E., Morata, L., Martínez-Pastor, J. C., Angulo, S., Combalia, A., Bori, G., & Soriano, A. (2018). Prosthetic joint infection due to *Pseudomonas aeruginosa*: Outcome of a difficult-to-treat infection with limited treatment options. *Journal of Clinical Medicine*, 7(3), 67.
- Tuon, F. F., Cieslinski, J., Ono, A. F. M., Goto, F. L., Machinski, J. M., Mantovani, L. K., Kosop, L. R., Namba, M. S., & Rocha, J. L. (2019). Microbiological profile and susceptibility pattern of surgical site infections related to orthopaedic trauma. *International Orthopaedics*, 43(6), 1309-1313. https://doi.org/10.1007/s00264-018-4076-7
- World Health Organization. (2017). Prioritization of pathogens to guide discovery, research and development of new antibiotics for drug-resistant bacterial infections including tuberculosis. Geneva: World Health Organization. (WHO/EMP/IAU/2017.12).
- World Health Organization. (2018). Infection control. Retrieved from <u>https://apps.who.int/iris/bitstream/handle/10665/2773</u> 99/9789241550475-eng.pdf?ua=1.
- Wu, W., Jin, Y., Bai, F., & Jin, S. (2015). Pseudomonas aeruginosa. In Molecular Medical Microbiology (pp. 753-767). Elsevier.
- Xu, Y., Chen, W., You, C., & Liu, Z. (2017). Development of a multiplex PCR assay for detection of *Pseudomonas fluorescens* with biofilm formation ability. *Journal of Food* Science, 82(10), 2337-2342. <u>https://doi.org/10.1111/1750-3841.13845</u>
- Yasidi, B. M., Denue, B. A., Onah, J. O., Jibrin, Y. B., Umar, H. M., Gabchiya, N. M., ... & Okon, K. O. (2015). Retrospective analysis of bacterial pathogens isolated from wound infections at a tertiary hospital in Nguru, Yobe State, Nigeria. *American Journal of Biomedical* and Life Sciences, 3(1), 1-6. https://doi.org/10.11648/j.asjbls.20150301.11